



March 2014 Volume 89 Number 3

PROCEEDINGS

Stethoscopes and Health Care—Associated Infection

ver the past 30 years we have come to fully appreciate the enormous potential for person-to-person spread of virulent nosocomial pathogens (eg, methicillinresistant Staphylococcus aureus [MRSA], vancomycin-resistant enterococcus [VRE], multidrug-resistant [MDR] gram-negative bacilli and Clostridium difficile, viruses such as influenza A, respiratory syncytial virus, and norovirus, and even Candida species) in the health care setting, with devastating infection being the most feared iatrogenic consequence and one of the greatest threats to hospital safety. 1,2 It has long been accepted that the major reservoir of nosocomial infection is infected or colonized patients and the major mode of transmission is the transient carriage of nosocomial pathogens on the hands of noncolonized health care workers having direct physical contact with patients.³ Hand hygiene before and after direct patient contact-now most often with a waterless alcohol gel or hand rub—has become an uncompromising expectation for modern-day health care workers.

Although it had long been held that microorganisms in the inanimate hospital environment do not play a significant role in the acquisition of nosocomial infection,⁵ it has become evident in recent years that surfaces in hospitals touched by patients or health care workers readily become contaminated by "environmental pathogens," such as MRSA, VRE, Acinetobacter baumanii, C difficile, respiratory syncytial virus, and norovirus, which collectively have a unique capacity to survive dessiccation in a viable, transmissible form for days to months. Compelling epidemiologic data indicate that contamination of inanimate surfaces in hospitals is an important reservoir of these pathogens and has driven a move toward more comprehensive surface decontamination

with bleach solutions, ultraviolet light, or aerosolization of hydrogen peroxide or peroxacetic acid.⁶

Auscultation of the heart, lungs, abdomen, and major arteries with a stethoscope has long been considered an integral part of the physical examination, and most health care providers prefer to use their own stethoscope. It has long been known that the diaphragms and bells of stethoscopes randomly sampled in a health care setting, such as a hospital, are almost universally contaminated by potential nosocomial pathogens, 7-18 most often staphylococci— MRSA up to 32% of the time 18 —but also Cdifficile, 17 resistant gram-negative bacilli, and even viruses, 19 and studies have shown that stethoscope contamination by these microorganisms is commonly acquired from colonized or infected patients. 9,10

In this issue of Mayo Clinic Proceedings, Longtin et al²⁰ report an innovative study of ungloved physicians who auscultated MRSAcolonized patients with presterilized stethoscopes, showing that the fingertips of the examiners or the diaphragms of their stethoscopes acquired MRSA contamination during 76% of the examinations. They found a powerful correlation between counts on examiners' hands and the quantitative level of contamination of the stethoscope with each examination, both for total bacterial counts and for MRSA. The efficiency of transmission of MRSA from the trunk of colonized or infected patients to the hands of health care workers and their stethoscopes rigorously documented in this unique real-life study is almost staggering. One can ask, why are we all not MRSA carriers?

Given that microorganisms on contaminated stethoscopes are readily transmitted

See also page 291

back to the surfaces they touch 7,9,10,12,14 and must be considered a preventable source of nosocomial colonization (and subsequent nosocomial infection) of patients, this mode of transmission would seem no less important than the uncleansed hands of health care workers. Although a number of studies have microbiologically and epidemiologically implicated electronic thermometers in the genesis of nosocomial outbreaks, 21-23 only a single report has linked the contamination of stethoscopes to infections in patients in an outbreak in a neonatal intensive care unit (ICU) epidemiologically as well as microbiologically²⁴ and no published study has made an unequivocal association with endemic health care—associated infections. However, if hand hygiene is considered an essential infection control measure to help prevent the spread of pathogens both in the health care setting and in the community, 4 it seems only logical that measures to minimize the accumulation of potential nosocomial pathogens on stethoscopes are needed to prevent transmission to vulnerable patients.

Studies have shown that wiping the head of a stethoscope with a 70% alcohol pledget ^{7-13,17} or wiping it with the antiseptic used for hand hygiene ^{7-9,16} or a hospital surface disinfectant ⁷⁻⁹ greatly reduces—usually eliminates—the bioburden of aerobic bacterial contamination. Moreover, the personal stethoscopes of health care workers who practice regular decontamination have been found to be less likely to be contaminated by MRSA and other MDR pathogens. ^{11-14,16} As such, health care workers should be expected to routinely decontaminate the head of their personal stethoscope between patients, logically when they do postexamination hand hygiene.

Unfortunately, the efficacy of these simple approaches to on-site decontamination of stethoscopes for removing *C difficile* spores or viruses that can also be present is unknown. It has long been accepted that dedicated stethoscopes—used only on the isolated patient and sent to Central Supply for decontamination with ethylene oxide gas when the patient is transferred or discharged—are an integral feature of barrier isolation to prevent the spread of microorganisms known to be spread by direct physical contact, such as

MRSA, VRE, MDR gram-negative bacilli, and C difficile but also all the respiratory and enteric viruses, and enteric parasites such as Giardia lamblia and Cryptosporidium species.²⁵ However, it has also become clear that for every patient known to be colonized or infected by an MDR nosocomial pathogen because of a positive clinical culture or C difficile polymerase chain reaction test, there are many more patients on that same patient care unit with undetected colonization—patients who pose a greater risk of spreading these microorganisms than patients known to be colonized or infected and in isolation.²⁶ This fact has formed the basis for "search and destroy" strategies for preventing MRSA and VRE infection, screening newly admitted patients for carriage to determine the need for barrier isolation and decolonization, 27,28 and most recently, putatively more cost-effective and broadly effective preventive strategies in the ICU, bypassing screening and subjecting all patients in the ICU to daily chlorhexidine bathing, ^{29,30} with or without the use of nasal mupirocin.31

Studies showing that neckties and clothing readily become contaminated by nosocomial pathogens such as MRSA or *C difficile* have driven a new policy in UK National Health Service hospitals forbidding neckties and jackets and mandating hospital-provided reprocessable overgarments for health care workers involved in direct patient care. Notwithstanding a recent multicenter trial that showed only moderate benefit, 33 preemptive barrier isolation of all high-risk patients, with dedicated stethoscopes, to prevent the spread of nosocomial pathogens has shown efficacy 34 and is practiced in many ICUs around the world.

In sum, I believe that it is now time for the use of dedicated stethoscopes with *all* ICU patients and a case can be made for all hospitalized patients. The complaint that the cheap stethoscopes many hospitals purchase for isolation rooms are barely functional can be obviated by purchasing higher quality stethoscopes in bulk with a unique and garish pattern on the tubing (eg, iridescent orange or striped) to deter theft, the major impediment to hospitals purchasing more expensive, high-quality institutional stethoscopes.

Promising advances in antiseptic surface technology to prevent surface microbial contamination^{35,36} may allow a return to the routine use of personal stethoscopes in the future.

Dennis G. Maki, MD

Madison WI

Divisions of Infectious Disease and Pulmonary/ Critical Care Medicine Department of Medicine University of Wisconsin School of Medicine and Public Health

Correspondence: Address to Dennis G. Maki, MD, Divisions of Infectious Disease and Pulmonary/Critical Care Medicine, Department of Medicine, University of Wisconsin School of Medicine and Public Health, 5227 UWMF Centennial Building, 1685 Highland Avenue, Madison, WI 53705-2281 (dgmaki@medicine.wisc.edu).

REFERENCES

- Maki DG, Tsigrelis C. Nosocomial infection in the intensive care unit. In: Parrillo JE, Dellinger RP, eds. Critical Care Medicine. Principles of Diagnosis and Management. (4th ed.). Philadelphia, PA: Mosby; 2014:825-869.
- Rosenthal VD, Bijie H, Maki DG, et al. International Nosocomial Infection Control Consortium (INICC) report, data summary of 36 countries, for 2004-2009. Am J Infect Control. 2012; 40(5):396-407.
- Maki DG. Control of colonization and transmission of pathogenic bacteria in the hospital. Ann Intern Med. 1978;89(5, Pt 2 Suppl):777-780.
- 4. Boyce JM, Pittet D; Healthcare Infection Control Practices Advisory Committee; HICPAC/SHEA/APIC/IDSA Hand Hygiene Task Force. Guideline for Hand Hygiene in Healthcare Settings: Recommendations of the Healthcare Infection Control Practices Advisory Committee and the HIPAC/SHEA/ APIC/IDSA Hand Hygiene Task Force. Am J Infect Control. 2002;30(8):S1-S46.
- Maki DG, Alvarado CJ, Hassemer CA, Zilz MA. Relation of the inanimate hospital environment to endemic nosocomial infection. N Engl J Med. 1982;307(25):1562-1566.
- Weber DJ, Rutala WA, Miller MB, Huslage K, Sickbert-Bennett E. Role of hospital surfaces in the transmission of emerging health care-associated pathogens: norovirus, Clostridium difficile, and Acinetobacter species. Am J Infect Control. 2010;38(5, Suppl 1):S25-S33.
- Jones JS, Hoerle D, Riekse R. Stethoscopes: a potential vector of infection? Ann Emerg Med. 1995;26(3):296-299.
- Marinella MA, Pierson C, Chenoweth C. The stethoscope. A potential source of nosocomial infection? Arch Intern Med. 1997;157(7):786-790.
- Bernard L, Kereveur A, Durand D, et al. Bacterial contamination of hospital physicians' stethoscopes. *Infect Control Hosp Epidemiol*. 1999;20(9):626-628.
- Zachary KC, Bayne PS, Morrison VJ, Ford DS, Silver LC, Hooper DC. Contamination of gowns, gloves, and stethoscopes with vancomycin-resistant enterococci. *Infect Control Hosp Epidemiol.* 2001;22(9):560-564.
- Parmar RC, Valvi CC, Sira P, Kamat JR. A prospective, randomised, double-blind study of comparative efficacy of immediate versus daily cleaning of stethoscope using 66% ethyl alcohol. Indian J Med Sci. 2004;58(10):423-430.

- Bandi S, Uddin L, Milward K, Aliyu S, Makwana N. How clean are our stethoscopes and do we need to clean them? J Infect. 2008;57(4):355-356.
- Fenelon L, Holcroft L, Waters N. Contamination of stethoscopes with MRSA and current disinfection practices. J Hosp Infect. 2009;71(4):376-378.
- Uneke CJ, Ogbonna A, Oyibo PG, Ekuma U. Bacteriological assessment of stethoscopes used by medical students in Nigeria: implications for nosocomial infection control. World Health Popul. 2008;10(4):53-61.
- Schroeder A, Schroeder MA, D'Amico F. What's growing on your stethoscope? (And what you can do about it). J Fam Pract. 2009;58(8):404-409.
- Uneke CJ, Ogbonna A, Oyibo PG, Onu CM. Bacterial contamination of stethoscopes used by health workers: public health implications. J Infect Dev Ctries. 2010;4(7):436-441.
- Vajravelu RK, Guerrero DM, Jury LA, Donskey CJ. Evaluation of stethoscopes as vectors of Clostridium difficile and methicillinresistant Staphylococcus aureus. Infect Control Hosp Epidemiol. 2012;33(1):96-98.
- Merlin MA, Wong ML, Pryor PW, et al. Prevalence of methicillin-resistant Staphylococcus aureus on the stethoscopes of emergency medical services providers. Prehosp Emerg Care. 2009;13(1):71-74.
- Blydt-Hansen T, Subbarao K, Quennec P, McDonald J. Recovery of respiratory syncytial virus from stethoscopes by conventional viral culture and polymerase chain reaction. *Pediatr Infect Dis J.* 1999;18(2):164-165.
- Longtin Y, Schneider A, Tschopp A, et al. Contamination of stethoscopes and physicians' hands following a physical examination. Mayo Clin Proc. 2014;89(3):291-299.
- Livomese LL Jr, Dias S, Samel C, et al. Hospital-acquired infection with vancomycin-resistant Enterococcus faecium transmitted by electronic thermometers. Ann Intern Med. 1992; 117(2):112-116.
- van den Berg RW, Claahsen HL, Niessen M, Muytjens HL, Liem K, Voss A. Enterobacter cloacae outbreak in the NICU related to disinfected thermometers. Hosp Infect. 2000;45(1): 29-34.
- Dijk Y, Bik EM, Hochstenbach-Vernooij S, et al. Management of an outbreak of Enterobacter cloacae in a neonatal unit using simple preventive measures. J Hosp Infect. 2002; 51(1):21-26.
- 24. Hoşbul T, Ozyurt M, Karademir F, Süleymanoğlu S, Haznedaroğlu T. Investigation of a nosocomial outbreak caused by ESBL positive Klebsiella pneumoniae in neonatal intensive care unit by AP-PCR. Mikrobiyol Bul. 2012;46(1): 101-105.
- Siegel JD, Rhinehart E, Jackson M, Chiarello L; Health Care Infection Control Practices Advisory Committee. 2007 Guideline for Isolation Precautions: Preventing Transmission of Infectious Agents in Health Care Settings. Am J Infect Control. 2007; 35(10, Suppl 2):S65-S164.
- Safdar N, Maki DG. The commonality of risk factors for nosocomial colonization and infection with antimicrobial-resistant Staphylococcus aureus, enterococcus, gram-negative bacilli, Clostridium difficile, and Candida. Ann Intern Med. 2002; 136(11):834-844.
- 27. Vos MC, Behrendt MD, Melles DC, et al. 5 years of experience implementing a methicillin-resistant Staphylococcus aureus search and destroy policy at the largest university medical center in the Netherlands. Infect Control Hosp Epidemiol. 2009; 30(10):977-984.
- 28. Jain R, Kralovic SM, Evans ME, et al. Veterans Affairs initiative to prevent methicillin-resistant *Staphylococcus aureus* infections. *N Engl | Med.* 2011;364(15):1419-1430.
- Bleasdale SC, Trick WE, Gonzalez IM, Lyles RD, Hayden MK, Weinstein RA. Effectiveness of chlorhexidine bathing to reduce catheter-associated bloodstream infections in medical

- intensive care unit patients. Arch Intern Med. 2007;167(19): 2073-2079.
- **30.** Climo MW, Yokoe DS, Warren DK, et al. Effect of daily chlor-hexidine bathing on hospital-acquired infection. *N Engl J Med.* 2013;368(6):533-542.
- Huang SS, Septimus E, Kleinman K, et al; CDC Prevention Epicenters Program; AHRQ DECIDE Network and Healthcare-Associated Infections Program. Targeted versus universal decolonization to prevent ICU infection. N Engl J Med. 2013; 368(24):2255-2265.
- 32. General Health Protection, Department of Health. Health Act 2006. Code of Practice for the Prevention and Control of Healthcare Associated Infections. Uniforms and Workwear. An Evidence Base for Developing Local Policy. London: Department of Health; 2006.
- Harris AD, Pineles L, Belton B, et al. Universal glove and gown use and acquisition of antibiotic-resistant bacteria in the ICU: a randomized trial. JAMA. 2013;310(15):1571-1580.
- Klein BS, Perloff WH, Maki DG. Reduction of nosocomial infection during pediatric intensive care by protective isolation. N Engl J Med. 1989;320(26):1714-1721.
- Karpanen TJ, Casey AL, Lambert PA, et al. The antimicrobial efficacy of copper alloy furnishing in the clinical environment: a crossover study. Infect Control Hosp Epidemiol. 2012;33(1): 3-9
- Bazaka K, Jacob MV, Crawford RJ, Ivanova EP. Efficient surface modification of biomaterial to prevent biofilm formation and the attachment of microorganisms. Appl Microbiol Biotechnol. 2012;95(2):299-311.